

This study highlights one of the most significant and recurring problems in ancient DNA research: that of authentication of results. While many labs are now careful to use strict experimental controls (as outlined in Cooper and Poinar⁴), the recommendation of phylogenetic verification of results continues to be ignored, despite the relative simplicity and lack of expense of such tests, particularly in comparison to experimental procedures. It should be noted that many of the most embarrassing mistakes in the ancient DNA literature (including the incorrect report of dinosaur DNA in the early 1990s⁵) could have been avoided with a simple phylogenetic test.

Conflict of interest: No conflict of interest to declare.

References

1. Cunha BA. The cause of the plague of Athens: plague, typhoid, typhus, smallpox or measles? *Infect Dis Clin N Am* 2004;**18**: 29–43.
2. Papagrigorakis MJ, Yapijakis C, Synodinos PN, Baziotopoulou-Valavani E. DNA examination of ancient dental pulp incriminates typhoid fever as a probable cause of the Plague of Athens. *Int J Infect Dis* 2006;**10**:206–14.
3. Ochman H, Elwyn S, Moran NA. Calibrating bacterial evolution. *Proc Natl Acad Sci USA* 1999;**96**:12638–43.
4. Cooper A, Poinar HN. Ancient DNA: do it right or not at all. *Science* 2000;**289**:1139.
5. Woodward SR, Weyland NJ, Bunnell M. DNA sequence from Cretaceous Period bone fragments. *Science* 2004;**266**:1229–32.
6. Swofford DL. *PAUP* Phylogenetic methods using parsimony (and other methods)*. Sunderland, MA, USA: Sinauer; 1999.

Beth Shapiro*

Andrew Rambaut

Henry Wellcome Ancient Biomolecules Centre,
Department of Zoology, University of Oxford,
South Parks Road, Oxford OX1 3PS, UK

M. Thomas P. Gilbert

Ancient DNA and Evolution,
Niels Bohr and Biological Institutes,
University of Copenhagen,
DK-2100 Copenhagen Ø, Denmark

*Corresponding author. Tel.: +44 1865 281248;
fax: +44 1865 271249

E-mail address: beth.shapiro@zoo.ox.ac.uk
(B. Shapiro)

Corresponding Editor: Jonathan Cohen, Brighton, UK

31 January 2006

doi:10.1016/j.ijid.2006.02.006

Insufficient phylogenetic analysis may not exclude candidacy of typhoid fever as a probable cause of the Plague of Athens (reply to Shapiro et al.)

The cause of the Plague of Athens (430–426 BC) has been debated among scientists, who have relied exclusively on Thucydides' historical narrations¹ to introduce several possible diagnoses.^{2,3} The application of DNA analysis on skeletal remains taken from the Kerameikos mass grave⁴ has been acknowledged as an ideal material that might provide clues for a definite evidence-based diagnosis of the epidemic.⁵ Following a research methodology of proven accuracy and validity ('suicide' PCR),⁶ it was shown by analysis of three genes (*osmC*, *clyA*, *narG*) that an ancient strain of *Salmonella enterica* serovar Typhi was present in the investigated dental pulp material of three putative victims of the plague, thus incriminating typhoid fever as a probable cause.⁷

Despite this evidence-documented approach, Shapiro et al.⁸ have argued against the validity of these results. Through the application of a simple phylogenetic analysis of the published sequence of one gene (*narG*), Shapiro et al. concluded that, although the sequenced ancient DNA quite possibly corresponds to a *Salmonella* strain, this might not be typhoid. Instead, they argue that this sequence is more closely matched to other *Salmonella* species such as *S. bongori*, *S. arizonae*, and *S. diarizonae* rather than *S. enterica* Typhi. The authors base this assumption on an inferred evolutionary timescale of *Salmonella* and other related bacterial species. Shapiro et al. eventually make the assumption that the identified sequence most probably represents a

modern and currently unknown free-living soil bacterium instead of an ancient one.

Against these arguments we must once again state the extreme preventative measures that were taken in our study⁷ to exclude any possibility of environmental contamination. These included the absence of the pathogens themselves or a previously attempted extraction or PCR amplification of the target DNA sequences in the implemented laboratories, and also the 'suicide' PCR methodology that was followed, which excluded positive controls from this study. In addition, since possible environmental contamination is a major problem with ancient DNA studies as Shapiro et al. suggest, soil wash was actually used as a negative control in addition to DNA extracts from modern teeth. As is clearly stated in the original publication of our results, no product was yielded following the application of the same primers under the same laboratory conditions on the negative controls as well as on the soil sample washed off the ancient teeth,⁷ thus excluding the possibility of any contamination of the investigated ancient material. Besides, *Salmonella* species do not survive for long in soil, which is typically regarded as a transitional environment for this pathogen prior to its infecting a host.⁹ Even if the soil of the mass grave was indeed contaminated by a modern *Salmonella* strain, such as a close relative of *S. bongori*, *S. arizonae*, and *S. diarizonae* (which are naturally found in reptiles), as suggested by Shapiro et al.,⁸ it would not be possible for any of these pathogens to survive during the long storage of the skeletal material and not be identified in the subsequently conducted investigation of the soil wash.

On the other hand, the application of phylogenetic models, as suggested by Shapiro et al.,⁸ undoubtedly constitutes a powerful tool for the introduction of theoretical assump-

tions and hypotheses where clear-cut evidence is not available. Nevertheless, it has been supported that there is no 'golden rule' to fit all cases of phylogenetic analysis, whereas a minimum of 6–10 comparable gene sequences are needed to permit a complete phylogenetic study.¹⁰ In contrast to these guidelines, the use of only one gene sequence for the application of phylogenetic analysis in bacteria, in which lateral gene transfer occurs widely between species,¹¹ might yield misleading results. Such seems to be the case for Shapiro et al.'s approach⁸ that applied their proposed model on only one gene sequence. Besides, while there have been some reports of molecular data, which are useful for construction of accepted phylogenetic trees of *Salmonella* species,^{12,13} other reports yield conflicting results. For example, the evolutionary relationships of *Salmonella* species based on synonymous site variation in the *fimA*, *fimI*, and *fimZ* genes, which belong in the same operon, result in diverse phylogenetic trees, possibly because of variable horizontal transmission of all or part of the same gene cluster.¹⁴ Another phylogenetic study of three genes placed most studied *Salmonella* species in monophyletic lineages, but one serotype was placed into two statistically well supported separate lineages which contained either bovine only or avian only isolates.¹⁵

In brief we doubt Shapiro et al.'s assumptions since:

- Environmental contamination of the investigated ancient material or its surrounding soil was specifically excluded by the precise methodology that was followed in our research study.
- A phylogenetic analysis of only one gene might be insufficient and/or misleading.
- The speculative results of the conducted phylogenetic analysis ("a modern and currently unknown free-living soil *Salmonella* species") cannot be more valid than the known clear evidence – *Salmonella* species do not survive for long in soil; dental pulp DNA sequences for two other genes have been detected, which not only are closest to *S. enterica* serovar Typhi, but one of these genes is lacking in its closest known relative, *S. typhimurium*.⁷

We agree with Shapiro et al.⁸ that there is not yet definite proof that the cause of the Plague of Athens was typhoid fever. The concurrent presence of a plurality of infectious diseases in besieged Athens of 430–426 BC was not excluded in the first place⁷ allowing for the variable clinical manifestation of the epidemic as reported by Thucydides.³

Conflict of interest: No conflict of interest to declare.

References

1. Thucydides. History of the Peloponnesian War. Books I & II, §2.47–55. London, UK: Harvard University Press; 1919. p. 341–59.
2. Durack DT, Littman RJ, Benitez RM, Mackowiak PA. Hellenic holocaust: a historical clinico-pathologic conference. *Am J Med* 2000;**109**:391–7.
3. Cunha BA. The cause of the plague of Athens: plague, typhoid, typhus, smallpox or measles? *Infect Dis Clin N Am* 2004;**18**:29–43.
4. Baziotopoulou-Valavani EA. Mass burial from the cemetery of Kerameikos. In: Stamatopoulou M, Yeroulanou M, editors. *Excavating classical culture. Recent archaeological discoveries in Greece. Studies in classical archaeology I. BAR International Series 1031*. Oxford, UK: Archaeopress; 2002. p. 187–201.
5. Holden C. (Random samples) Athenian plague probe. *Science* 1996;**274**:1307.
6. Raoult D, Aboudharam G, Crubezy E, Larroy Ludes B, Drancourt M. Molecular identification by suicide PCR of *Yersinia pestis* as the agent of medieval black death. *Proc Natl Acad Sci USA* 2000;**23**:12800–3.
7. Papagrigorakis MJ, Yapijakis C, Synodinos PN, Baziotopoulou-Valavani E. DNA examination of ancient dental pulp incriminates typhoid fever as a probable cause of the Plague of Athens. *Int J Infect Dis* 2006;**10**:206–14.
8. Shapiro B, Rambault A, Gilbert TP. No proof that typhoid caused the Plague of Athens. *Int J Infect Dis* 2006; doi:10.1016/j.ijid.2006.02.006.
9. Winfield MD, Groisman EA. Role of nonhost environments in the lifestyles of *Salmonella* and *Escherichia coli*. *Appl Environ Microbiol* 2003;**69**:3687–94.
10. Cummings MP, Meyer A. Magic bullets and golden rules: data sampling in molecular phylogenetics. *Zoology* 2005;**108**:329–36.
11. Hoshino T, Fujiwara T, Kilian M. Use of phylogenetic and phenotypic analyses to identify nonhemolytic streptococci isolated from bacteremic patients. *J Clin Microbiol* 2005;**43**:6073–85.
12. Kanluk NA, Monteiro MA, Parker CT, Whitfield C. Molecular diversity of the genetic loci responsible for lipopolysaccharide core oligosaccharide assembly within the genus *Salmonella*. *Mol Microbiol* 2002;**46**:1305–18.
13. Chan K, Baker S, Kim CC, Detweiler CS, Dougan G, Falkow S. Genomic comparison of *Salmonella enterica* serovars and *Salmonella bongori* by use of an *S. enterica* serovar Typhimurium DNA microarray. *J Bacteriol* 2003;**185**:553–63.
14. Boyd EF, Hartl DL. Analysis of the type 1 pilin gene cluster *fim* in *Salmonella*: its distinct evolutionary histories in the 5' and 3' regions. *J Bacteriol* 1999;**181**:1301–8.
15. Sukhnanand S, Alcaine S, Warnick LD, Su WL, Hof J, Craver MPJ, et al. DNA sequence-based subtyping and evolutionary analysis of selected *Salmonella enterica* serotypes. *J Clin Microbiol* 2005;**43**:3688–98.

Manolis J. Papagrigorakis^{a,d,*}
 Christos Yapijakis^a
 Philippos N. Synodinos^b
 Effie Baziotopoulou-Valavani^c

^aDepartment of Orthodontics, Dental School,
 University of Athens, 2 Thivon str.,
 11527 Goudi/Athens, Greece

^bDepartment of Neurology, University of Athens Medical School,
 Athens, Greece

^cThird Ephorate of Prehistoric and Classical Antiquities,
 Greek Ministry of Culture, Athens, Greece

^dMuseum of Craniofacial Evolution and Dental History,
 Dental School, University of Athens,
 Goudi/Athens, Greece

*Corresponding author. Tel.: +30 2107461229;
 fax: +30 2106106244

E-mail address: demon@otenet.gr (M. J. Papagrigorakis)

Corresponding Editor: Jonathan Cohen, Brighton, UK

22 February 2006

doi:10.1016/j.ijid.2006.02.005